

## Test Description

The MolQ Liquid Precision Panel includes 50 genes, involving hotspot regions and 3159 unique variants, applicable to a wide range of tumor types for detection of SNV (single and multiple nucleotide variation), Insertion-Deletion, Copy Number Variation (CNV), and gene Fusions. Fusion and splice variants are detected in RNA.

## Patient Demographic

**Name:** Ms. Megha Kalyan  
**Sex:** Female  
**Date of Birth/Age:** 42 years  
**Disease:** Lung Adenocarcinoma

## Clinician

**Clinician Name:** Dr Amit Verma  
**Medical Facility:** Dr AV Institute of Personalized Cancer Therapy and Research  
**Pathologist:** Not Provided

## Specimen

**Booking ID:** 012406220051  
**Sample Type:** Blood  
**Tumor Content Percentage:** NA  
**Date of Collection:** 22-06-2024  
**Date of Booking:** 22-06-2024

## CLINICAL SYNOPSIS

Megha Kalyan, is a known case of lung adenocarcinoma. She has been evaluated for pathogenic variations in the genes listed in Appendix 2.

## RESULT SUMMARY

**Variants detected as per NCCN Guidelines:** Clinically relevant *EGFR* exon 19 deletion *EGFR* (p.Glu746\_Ala750del, VAF= 0.10%) mutation is detected.

### Other variants detected:

Not Applicable

Note: The sequencing was performed on 26.6 ng of cfTNA in the given specimen. Average Base Coverage Depth achieved was 57370X in this sample.

*"Important Disclaimer: Note: In the present specimen, the given variant is below our reporting threshold for VAF%. However, this variant was present in the previous reports of solid and liquid analyzed from the patient specimen of tissue and blood in May-June 2023."*

## RESULTS

**Variant in *EGFR* gene is detected.**

Gene/ Transcript (Locus)	Variant ID	Variant	Exon	Coverage	Allele Frequency	Variant Effect	*Relevant Therapies (In this cancer type)	(In other cancer type)	Tier <sup>1</sup>
<i>EGFR</i> NM_005228.5 (chr7:55242465)	COSM6225	c.2236_2250d eIGAAT TAAGAGAAGC A (p.Glu746_Ala 750del)	19	1945	0.01%	Non frame shift Deletion	Afatinib <sup>i,ii</sup> bevacizumab* + erlotinib <sup>ii</sup> dacomitinib <sup>i,ii</sup> erlotinib <sup>i,ii</sup> erlotinib + ramucirumab <sup>i,ii</sup> gefitinib* <sup>i,ii</sup> osimertinib <sup>i,ii</sup> osimertinib + chemotherapy <sup>i</sup> gefitinib + chemotherapy amivantamab + chemotherapy atezolizumab + bevacizumab + chemotherapy	None	Ia

\* Public data sources included in relevant therapies: FDA<sup>1</sup>, NCCN, EMA<sup>2</sup>, ESMO

<sup>1</sup>Li et al. Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn. 2017 Jan;19(1):4-23.

## RELEVANT NON-SMALL CELL LUNG CANCER FINDINGS

Gene	Findings	Gene	Findings	Gene	Findings
<i>ALK</i>	None detected	<i>KRAS</i>	None detected	<i>NTRK3</i>	None detected
<i>BRAF</i>	None detected	<i>MET</i>	None detected	<i>RET</i>	None detected
<i>EGFR</i>	Exon 19 deletion	<i>NTRK1</i>	None detected	<i>ROS1</i>	None detected
<i>ERBB2</i>	None detected	<i>NTRK2</i>	None detected		

## CLINICAL CORRELATION AND VARIANT INTERPRETATION

### ***EGFR* p.Glu746\_Ala750del (Exon 19 deletion) Coverage 1945**

**Gene description:** The *EGFR* gene encodes the epidermal growth factor receptor (EGFR) tyrosine kinase, a member of the ERBB/human epidermal growth factor receptor (HER) family. In addition to EGFR/ERBB1/HER1, other members of the ERBB/HER family include ERBB2/HER2, ERBB3/HER3, and ERBB4/HER4<sup>1</sup>. EGFR ligand induced dimerization results in kinase activation and leads to stimulation of oncogenic signaling pathways including the PI3K/AKT/MTOR and RAS/RAF/MEK/ERK pathways. Activation of these pathways promote cell proliferation, differentiation, and survival<sup>2,3</sup>.

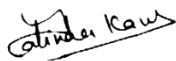
**Alterations and prevalence:** Recurrent somatic mutations in the tyrosine kinase domain (TKD) of *EGFR* are observed in approximately 10-20% of lung adenocarcinoma, and at higher frequencies in never-smoker, female, and Asian populations<sup>4-7</sup>. The most common mutations occur near the ATP-binding pocket of the TKD and include short in-frame deletions in exon 19 (*EGFR* exon 19 deletion) and the L858R amino acid substitution in exon 21<sup>8</sup>. These mutations constitutively activate EGFR resulting in downstream signaling, and represent 80% of the *EGFR* mutations observed in lung cancer. A second group of less prevalent activating mutations include E709K, G719X, S768I, L861Q, and short in-frame insertion mutations in exon 20<sup>9-12</sup>. *EGFR* activating mutations in lung cancer tend to be mutually exclusive to *KRAS* activating mutations<sup>13</sup>. In contrast, a different set of recurrent activating *EGFR* mutations in the extracellular domain include R108K, A289V and G598V and are primarily observed in glioblastoma<sup>8,14</sup>. Amplification of EGFR is observed in several cancer types including 30% of glioblastoma, 12% of esophageal cancer, 10% of head and neck cancer, 5% of bladder cancer, and 5% of lung squamous cell carcinoma<sup>4,5,7,14,15</sup>. Deletion of exons 2-7, encoding the extracellular domain of *EGFR* (EGFRvIII), results in overexpression of a ligand-independent constitutively active protein and is observed in approximately 30% of glioblastoma<sup>16-18</sup>.

**Potential relevance:** Approved first-generation EGFR tyrosine kinase inhibitors (TKIs) include erlotinib<sup>19</sup> (2004) and gefitinib<sup>20</sup> (2015), which block the activation of downstream signaling by reversible interaction with the ATP-binding site. Although initially approved for advanced lung cancer, the discovery that drug sensitivity was associated with exon 19 and exon 21 activating mutations allowed first-generation TKIs to become subsequently approved for front-line therapy in lung cancer tumors containing exon 19 or exon 21 activating mutations. Second-generation TKIs afatinib<sup>21</sup> (2013) and dacomitinib<sup>22</sup> (2018) bind EGFR and other ERBB/HER gene family members irreversibly and were subsequently approved. First- and second-generation TKIs afatinib, dacomitinib, erlotinib, and gefitinib are recommended for the treatment NSCLC harboring *EGFR* exon 19 insertions, exon 19 deletions, point mutations L861Q, L858R, S768I and codon 719 mutations, whereas most *EGFR* exon 20 insertions, except p.A763\_Y764insFQEA, confer resistance to the same therapies<sup>23-26</sup>. However, in 2021, the irreversible tyrosine kinase inhibitor, mobocertinib<sup>27</sup> was FDA approved for the treatment of NSCLC with *EGFR* exon 20 insertion mutations.

Additionally, in 2022, the FDA granted breakthrough therapy designation to the irreversible EGFR inhibitors, CLN-081 (TPC-064)<sup>28</sup> and sunvozertinib<sup>29</sup>, for locally advanced or metastatic non-small cell lung cancer harboring *EGFR* exon 20 insertion mutations. In lung cancer containing *EGFR* exon 19 or 21 activating mutations, treatment with TKIs is eventually associated with the emergence of drug resistance<sup>30</sup>. The primary resistance mutation that emerges following treatment with first-generation TKI is T790M, accounting for 50-60% of resistant cases<sup>8</sup>. Third generation TKIs were developed to maintain sensitivity in the presence of T790M. Osimertinib<sup>31</sup> (2015) is an irreversible inhibitor indicated for metastatic *EGFR* T790M positive lung cancer and for the first-line treatment of metastatic NSCLC containing *EGFR* exon 19 deletions or exon 21 L858R mutations. Like first-generation TKIs, treatment with osimertinib is associated with acquired resistance. In this case, resistance is associated with the C797S mutation and occurs in 22-44% of cases<sup>30</sup>. The T790M and C797S mutations may be each selected following sequential treatment with a first-generation TKI followed by a third-generation TKI or vice versa<sup>32</sup>. T790M and C797S can occur in either cis or trans allelic orientation<sup>32</sup>. If C797S is observed following progression after treatment with a third-generation TKI in the first-line setting, sensitivity may be retained to first-generation TKIs<sup>32</sup>. If C797S co-occurs in trans with T790M following sequential treatment with first- and third-generation TKIs, patients may exhibit sensitivity to combination first- and third-generation TKIs, but resistance to third-generation TKIs alone<sup>32,33</sup>. However, C797S occurring in cis conformation with T790M, confers resistance to first- and third-generation TKIs<sup>32</sup>. Fourth-generation TKIs are in development to overcome acquired C797S and T790M resistance mutations after osimertinib treatment. EGFR targeting antibodies including cetuximab (2004), panitumumab (2006), and necitumumab (2016) are under investigation in combination with EGFR-targeting TKIs for efficacy against *EGFR* mutations. The bispecific antibody, amivantamab<sup>34</sup>, targeting EGFR and MET was approved (2021) NSCLC tumors harboring *EGFR* exon 20 insertion mutations. In 2024, a CNS penetrating small molecule, ERAS-801<sup>35</sup> received fast track designation for the treatment of adult patients with *EGFR* altered glioblastoma. HLX-42<sup>36</sup>, an anti-EGFR-antibody-drug conjugate (ADC) consisting of an anti-EGFR monoclonal antibody conjugated with a novel high potency DNA topoisomerase I (topo I) inhibitor, received a fast-track designation (2024) for the treatment of patients with advanced or metastatic *EGFR*-mutated non-small cell lung cancer whose disease has progressed on a third-generation EGFR tyrosine kinase inhibitor. CPO301<sup>37</sup> received a fast-track designation (2023) from the FDA for EGFR mutations in patients with metastatic NSCLC who are relapsed/refractory or ineligible for EGFR targeting therapy such as 3rd-generation EGFR inhibitors including osimertinib. The Oncoprex immunogene therapy quaratusugene ozeplasmid<sup>38</sup> in combination with osimertinib received a fast-track designation from the FDA (2020) for NSCLC tumors harboring EGFR mutations that progressed on osimertinib alone. BDTX-189<sup>39</sup> was granted a fast-track designation (2020) for the treatment of solid tumors harboring an *EGFR* exon 20 insertion mutation.

## RECOMMENDATIONS

- Validation of the variant(s) by Sanger sequencing is recommended to rule out false positives.
- Genetic counselling is advised for interpretation on the consequences of the variant(s).
- If results obtained do not match the clinical findings, additional testing should be considered as per referring clinician's recommendations.



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## APPENDIX 1: TEST METHODOLOGY

### Method

Circulating cell-free total nucleic acid (cfTNA) were isolated from samples using the MagMAX Cell-Free Total Nucleic Acid Isolation Kit. Quantity and quality is checked by Qubit assay and Tape station, respectively. After quality check the isolated and purified sample was directly loaded on Ion Torrent Genexus Next Generation Sequencer and subjected to automated library preparation and template preparation followed by sequencing at an average depth of ~30000X.

It utilizes unique molecular tags to enable high sensitivity detection of variants. Analysis is done using Ion Torrent Reporter Software (version 6.6.2.1), the data is visualized on Integrative Genomics Viewer (IGV, version 5.01 (0)) and analyzed. The final report is generated using OncoPrint curated knowledgebase reporter and includes clinical trials information continuously being updated for the best of the patient management as per clinical guidelines.

### DISCLAIMER

- This report was generated using the materials and methods as recommended which required the use of quality reagents, protocols, instruments, software, databases and other items, some of which were provided or made accessible by third parties. A defect or malfunction in any such reagents, protocols, instruments, software, databases and/or other items may compromise the quality or accuracy of the report.
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- Medical knowledge and annotation are constantly updated and reflects the current knowledge at the time.
- Due to inherent technology limitations of the assay, not all bases of the exome can be covered by this test. Accordingly, variants in regions of insufficient coverage may not be identified and/or interpreted. Therefore, it is possible that certain variants are present in one or more of the genes analyzed, but have not been detected. The variants not detected by the assay that was performed may/ may not impact the phenotype.
- It is also possible that a pathogenic variant is present in a gene that was not selected for analysis and/or interpretation in cases where insufficient phenotypic information is available.
- The report shall be generated within turnaround time (TAT), however, such TAT may vary depending upon the complexity of test(s) requested. MolQ Laboratory under no circumstances will be liable for any delay beyond afore mentioned TAT.
- It is hereby clarified that the report(s) generated from the test(s) do not provide any diagnosis or opinion or recommends any cure in any manner. MolQ Laboratory hereby recommends the patient and/or the guardians of the patients, as the case may be, to take assistance of the clinician or a certified physician or doctor, to interpret the report(s) thus generated. MolQ Laboratory hereby disclaims all liability arising in connection with the report(s).
- In a very few cases genetic test may not show the correct results, e.g. because of the quality of the material provided to MolQ Laboratory. In case where any test provided by MolQ Laboratory fails for unforeseeable or unknown reasons that cannot be

influenced by MolQ Laboratory in advance, MolQ Laboratory shall not be responsible for the incomplete, potentially misleading or even wrong result of any testing if such could not be recognized by MolQ Laboratory in advance.

- A negative value in liquid biopsy does not mean true absence of mutation. It may not be detectable in the blood sample but may still be positive in tissue biopsy.
- This is a laboratory developed test and the development and the performance characteristics of this test was determined by reference laboratory as required by the CLIA 1988 regulations. The report, and the tests used to generate the Report have not been cleared or approved by the US Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. The test results have scientifically shown to be clinically useful.

## LIMITATIONS

- Variants with very low allele frequency (<0.5%) present in the given specimen or lower copy number variation might not be detected. Similarly fusion variants with less read may not be detected in liquid biopsy. Variant detection is also based on release of tumor cells or their fractions in the blood stream, it is affected by several factors.
- A negative report on liquid biopsy does not rule out the absence of variant.

**APPENDIX 2: GENE LIST**

DNA Hotspots					
<i>AKT1</i>	<i>AKT2</i>	<i>AKT3</i>	<i>ALK</i>	<i>AR</i>	<i>ARAF</i>
<i>BRAF</i>	<i>CDK4</i>	<i>CDKN2A</i>	<i>CHEK2</i>	<i>CTNNB1</i>	<i>EGFR</i>
<i>ERBB2</i>	<i>ERBB3</i>	<i>ERBB4</i>	<i>ESR1</i>	<i>FGFR1</i>	<i>FGFR2</i>
<i>FGFR3</i>	<i>FGFR4</i>	<i>FLT3</i>	<i>GNA11</i>	<i>GNAQ</i>	<i>GNAS</i>
<i>HRAS</i>	<i>IDH1</i>	<i>IDH2</i>	<i>KIT</i>	<i>KRAS</i>	<i>MAPK1</i>
<i>MAPK2</i>	<i>MET</i>	<i>MTOR</i>	<i>NRAS</i>	<i>NTRK1</i>	<i>NTRK2</i>
<i>NTRK3</i>	<i>PDGFRA</i>	<i>PIK3CA</i>	<i>PTEN</i>	<i>RAF1</i>	<i>RET</i>
<i>ROS1</i>	<i>SMO</i>	<i>TP53</i>			
CNVs					
<i>ALK</i>	<i>AR</i>	<i>CD274</i>	<i>CDKN2A</i>	<i>EGFR</i>	<i>ERBB2</i>
<i>ERBB3</i>	<i>FGFR1</i>	<i>FGFR2</i>	<i>FGFR3</i>	<i>KRAS</i>	<i>MET</i>
<i>PIK3CA</i>	<i>PTEN</i>				
Inter-genetic Fusions					
<i>ALK</i>	<i>BRAF</i>	<i>ESR1</i>	<i>FGFR1</i>	<i>FGFR2</i>	<i>FGFR3</i>
<i>MET</i>	<i>NRG1</i>	<i>NTRK1</i>	<i>NTRK2</i>	<i>NTRK3</i>	<i>NUTM1</i>
<i>RET</i>	<i>ROS1</i>	<i>RSPO2</i>	<i>RSPO3</i>		
Intra-genetic Fusions					
<i>AR</i>	<i>EGFR</i>	<i>MET</i>			