	Germline Cancer Predisposition-Additional
•	Family Member (Investigational) Testing

The MolQ Germline Cancer Predisposition-Additional Family

Member (Investigational) Testing analyse variant(s) observed in

other family members by targeted gene Sanger sequencing.

PATIENT	REPORT DATE	BOOKING ID
Ashok Sharma	23 May 2023	#012304120159

## Clinician

Clinician Name: Dr Amit Verma Medical Facility: Dr AV Institute of Personalized Therapy and Cancer Research (IPTCR) Pathologist: Not Provided

#### Specimen

Booking ID: 012304120159 Site: NA Sample Type: Blood Date of Collection:12-04-2023 Date of Booking:12-04-2023

# **CLINICAL SYNOPSIS**

**Patient Demographic** 

Name: Mr Ashok Sharma

**Disease**: Asymptomatic

Date of Birth/Age: 70 years

Sex: Male

**Test Description** 

The index patient, Mr. Anil Sharma (Sample ID: 7752274), is a case of metastatic, small cell, neuroendocrine carcinoma, prostate. He has a family history of prostate cancer with his brother diagnosed at the age of 48 years. He was found to harbor a heterozygous pathogenic variant, c.2317del in the *PALB2* gene. His sibling is being evaluated for the same variant.

### RESULTS

Variant is detected

Gene#	Location	Variant	Zygosity	Clinical condition of family member	Variation reported in family member*
PALB2	Exon 5	chr16:g.23629837del (GRCh38); c.2317del (HET); (p.Thr773LeufsTer78)	Heterozygous	Asymptomatic	Present
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\*The variant analysis in Sanger sequencing is based on the *PALB2* reference sequence ENST00000261584.8<sup>1</sup>. The exon number and nucleotide numbers will differ based on the reference file chosen and the database used.

### **CLINICAL CORRELATION AND VARIANT INTERPRETATION**

*Variant description:* A heterozygous single base pair deletion in exon 5 of the *PALB2* gene (**chr16:g.23629837del; c.2317del**) that results in a frameshift and premature truncation of the protein 78 amino acids downstream to codon 773 (**p.Thr773LeufsTer78**) was detected in the index patient (Sample ID: 7752274; Report Dated: 1st December 2022) by NGS and was further validated by Sanger sequencing.

The same variant is detected in heterozygous condition in the asymptomatic sibling of the index patient, Mr. Ashok Sharma (Figure 1).

The variant detected in the test and its significance needs to be carefully correlated with the clinical indications of the index patient.

### **RECOMMENDATIONS**

Genetic counselling is advised to discuss and interpret the significance of the results. Kindly email us at contact@molq.in for post-test counselling.

MolQ Laboratory (A Unit of Molecular Quest Healthcare Pvt. Ltd.)

Reference Laboratory: 28-29, Sector-18 (P) | Gurgaon, Haryana, 122015 | Phone 0124 - 4307906, Fax 0124 - 4278596 | Email: contact@molq.in





1. ENSEMBL: http://www.ensembl.org.

Figure 1: Sequence chromatogram and alignment to the reference sequence showing the variant in exon 5 of the *PALB2* gene (chr16:g.23629837del; c.2317del; p.Thr773LeufsTer78) detected in heterozygous condition in the sibling of the index patient, Mr. Ashok Sharma.

			A <mark>WTCCAAMRKTCCRGGMRCCCC</mark> AATTCGACAGTTCAGGCAGCCC:
2 fragment bases at consensus position 153	I30 TGAAAGACTCAGTCT	GTCTTGCCAGTGATACTAAAC	160 170 1 AATYCRACAGTYCAGGCASCCC
	PALB2	+++ ++ ; c.2317del ; chrl6:g.	+ • • +++ • + ++• .23629837del
<u>c r g r c r</u>	ΤΤ G C C A (	PALB2_e_05_7752274_s1_F_202 G_T_G_A_T_M_Y_W_A	A A M A W T C C

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Jatinder Kaur, PhD Head, Molecular Biology & Genomics

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Dr. Gulshan Yadav, MD Head, Pathology

# Germline Cancer Predisposition-Additional Family Member (Investigational) Testing

## **APPENDIX 1: TEST METHODOLOGY**

### **METHOD**

**Targeted Gene Sanger Sequencing**: Exon 5 of the *PALB2* gene was PCR-amplified and the product was sequenced using Sanger sequencing. In case of mosaicism in leucocytes, the detection limits of Sanger sequencing for presence of variant are  $\sim$ 20%. The sequence was aligned to available reference sequence ENST00000261584.8<sup>1</sup> to detect variant using variant analysis software programs. Variant classification follows the tenets of American College of Medical Genetics (ACMG) guidelines<sup>2</sup>.

### DISCLAIMER

- 1. This is a laboratory developed test and the development and the performance characteristics of this test was determined by the reference laboratory.
- 2. Please note that the tests are performed only after approval of referring/ ordering clinician/physician. Above recommendations /results should not be viewed as only source of information on which treatment or other clinical decisions are made. Clinical correlation is highly recommended.
- 3. The classification of variants of unknown significance can change over time and MolQ Laboratory cannot be held responsible for this. Please contact MolQ Laboratory later to inquire about any changes.
- 4. This sanger custom test is not a clinically validated assay for each, and every primer set. No form of test can guarantee 100% accuracy. This assay is no such exception, and it has some Inherent pitfalls.
- 5. The PCR based assay will not amplify genic regions outside primer binding site, may not detect the exact size for the deletion or duplication which is more than 50bp.
- 6. Though PCR is a highly sensitive and specific technique, performance may vary based on several factors including allelic dropout & preferential amplification (Chance phenomenon) causing a potential source of misdiagnosis for both dominant and recessive diseases. About 0.44% of total cases are susceptible to allele dropout phenomenon, which can lead to misdiagnosis<sup>3</sup>.
- 7. Testing of affected/carrier index/proband samples parallel with test samples is highly recommended to rule out false negative/positive results.
- 8. The accuracy of the results assumes that samples received were correctly identified, family relationships are true and clinical diagnosis of relatives is correct.
- 9. In a very few cases genetic tests may not show the correct results leading to false positives and negatives, e.g., because of the quality of the sample provided to MolQ Laboratory. In case where any test provided by MolQ Laboratory fails for unforeseeable or unknown reason that cannot be influenced by MolQ Laboratory in advance, MolQ Laboratory shall not be responsible for the incomplete, potentially misleading, or even wrong result of testing if such could not be recognized by MolQ Laboratory in advance.
- 10. Negative results do not negate the absence of mutations that are not covered by the test.
- 11. The report shall be generated within turnaround time (TAT), however, such TAT may vary depending upon the complexity of test(s) requested. MolQ Laboratory under no circumstances will be liable for any delay beyond afore mentioned TAT.
- 12. If results obtained do not match the clinical findings, additional testing should be considered as per the referring clinician's recommendations.
- 13. MolQ Laboratory hereby recommends the patients and/ or guardians of the patients, as the cause may be, to take assistance of the clinician or a certified physician or doctor, to interpret the report(s) thus generated. MolQ Laboratory hereby disclaims all liability arising in connection with the report (s).

### REFERENCES

- 1. ENSEMBL: http://www.ensembl.org.
- 2. Green RC et al. ACMG recommendations for reporting of incidental findings in clinical exome and genome sequencing. Genet Med. 2013, 15(7):565-74.
- 3. Blais J et al. Risk of Misdiagnosis Due to Allele Dropout and False-Positive PCR Artifacts in Molecular Diagnostics: Analysis of 30,769 Genotypes. J Mol Diagn. 2015, 17(5): 505-14.

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