

Prof. Dr. HEMANT MALHOTRA

MD, FRCP (London), MNAMS, FICP, FUICC, FIMSA
RMC Reg No. 619/800

Medical Oncology Specialization

Tata Memorial Hospital, Bombay
New York Hospital-Cornell Medical Center, New York
Sloan Kettering Cancer Center, New York

Senior Professor of Medicine &
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SMS Medical College Hospital, Jaipur - 302 004
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Fax 91-141-5105589
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Consultant Medical Oncologist & Hematologist
C-70, Ram Marg, Tilak Nagar, Jaipur - 302 004
Phone 91-141-2620500, 4004647
Fax 91-141-2622899
E-mail drmalhotrahemant@gmail.com

No myc

Dr. Pankaj Bainsi

20/10/2015

021 g/e fem ASD

36

N. P. Chandra
N. S. Chandra
N. S. Chandra

Dr. P. Chandra
Dr. P. Chandra
Dr. P. Chandra

15/03/2015

Dr. P. Chandra

400/48x 19/11/17 No myc

Dr. P. Chandra

12/3/15
021 mm x 4ub
12/3/15
11/12/15

Dr. P. Chandra

12/3/15 x 4ub
11/12/15

Dr. P. Chandra

26/01/16/ No myc

Dr. P. Chandra

at 12/3/15

Dr. P. Chandra

12/3/15

Dr. P. Chandra

12/3/15

Dr. P. Chandra

40/05/2016/ No myc

Dr. P. Chandra

13/06/2016/ No myc

Dr. P. Chandra

19/07/2016/ No myc

Dr. P. Chandra

12/08/16/ No myc

Dr. P. Chandra

21/09/16/ No myc

Dr. P. Chandra

12/10/16/ No myc

Dr. P. Chandra

17/11/16/ No myc

Dr. P. Chandra

22/12/16/ No myc

Dr. P. Chandra

26/01/17/ No myc

Dr. P. Chandra

29/02/17/ No myc

Dr. P. Chandra

Consultation by appointment only.
In case of emergency, please report immediately to the A&E department of the SMS Hospital, Jaipur.

01413727700

Prof. Dr. HEMANT MALHOTRA

MD, FRCP (London), MNAMS, FICP, FUICC, FIMSA
RMC Reg No : 619/8800.

Medical Oncology Specialization

Tata Memorial Hospital, Bombay
New York Hospital-Cornell Medical Center, New York
Sloan Kettering Cancer Center, New York

H-156
6-46

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Phone 91-141-2620600, 4004647
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E-mail drmalhotrahemant@gmail.com

cl. Jena
w. e. s. m.
ok. G. e. J. a. i. r. e. s.
P. a. i. l. i. t. + N. o. W. a. n. t. h. e.
S. t. a. m. e. l. k. a. n. d. +
H. - S. +
C. - c. l. e. m.
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s. p. F. e. n.

Dr. Parth Sarin 17/10/12
34

A. C. M. L. - C. P.
For LM / for GIPAP / G. linee

120/400.0/128.0
P. S. T. E. C. S. 204. M. O. 2. B. O. 4
B. C. C. G. M. C. C. S. M. M. C. I. Y. P. M. C. U. 2. B. O. 5
G. I. P. A. P. C. C. E. S. / F. G. W. / 12.10.12
F. m. H. y. W. / G. a. l. i. p. / B. o. m. b. e. y.
P. m. 02. B. U. 2.

C. G. linee 400 1.00 ✓
C. Arney-D 1.00 A. e.
C. Adesb (F 1.00)
C. B. a. e. d. a. e. P. B. 1.00 ✓
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07/11/2012 cl. Jena
04/12/2012 C. G. linee 400 / 4.1300m

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H. l. e. / A. t. h. o. r. m. m.

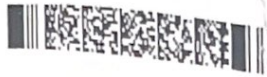
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24/12/2012
2. L. a. m. e. l. c. o. l. i. c. / 20.00 Dr. Vinay Kumar
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CODE : C000014810

PATIENT'S NAME AND ADDRESS :
DR. HAMENT MALHOTRA (DR)
RAJENDRA SINGH CIRCLE,

POUR 340001
RAJASTHAN INDIA
0141-3245299



SRL LIMITED
PRIME SQUARE BUILDING, PLOT NO 1, GAIWADI INDUSTRIAL
ESTATE, S.V. ROAD, GOREGAON (W)
Mumbai, 400062
MAHARASHTRA, INDIA
Tel : 022 6780 1177, 1-800-222-660, Fax : 022 - 67801212
CTN - U74899DL1995PLC070603
Email : srl.mumbai@srl.in

PATIENT NAME : PAPPU RAM SAINI

ACCESSION NO : 00020D047071

AGE : 36 Years

SEX : Male

PATIENT ID :

DRAWN : 14/04/2015 17:02

RECEIVED : 16/04/2015 03:26

DATE OF BIRTH :

REFERRING DOCTOR : DR. Hament Malhotra

REPORTED : 20/04/2015 15:28

Test Report Status **Final**

Results

CLIENT PATIENT ID : NSM000450/15

MOLECULAR BIOLOGY

BCR-ABL TRANSCRIPT QUANTIFICATION

BCR-ABL TRANSCRIPT QUANTIFICATION

METHOD : REAL TIME PCR

0.00

%

9829989993

Interpretation(s)

BCR-ABL TRANSCRIPT QUANTIFICATION-

Real Time Reverse Transcriptase Polymerase Chain Reaction

The BCR/ ABL gene rearrangement is found in 95% of patients with CML and 5% of children & 15% to 30% of adults with acute lymphoblastic leukemia(ALL). These cases are characterized by an aberrant gene rearrangement between the ABL protooncogene on chromosome 9 and the breakpoint cluster region (BCR) on chromosome 22.

Clinical Utility: Quantitative analysis of BCR/ABL transcript levels by Real Time PCR helps to monitor patients on therapy. Baseline and follow up assessments help to reliably identify the degree of molecular response, early impending relapse, and resistance to therapy. During early chronic phase, Major Molecular Response (MMR), which refers to a 3 fold reduction from a standard baseline value, is usually considered as a successful treatment outcome. The ultimate goal of treatment is to achieve Complete Molecular Response (CMR), which refers to not detectable (or 0%) BCR/ABL transcript levels by Real Time PCR.

Interpretation: BCR-ABL transcript quantification is expressed as a % of the control gene, i.e., ABL. The sample showing amplification curve for both BCR/ABL transcript and control gene are considered as positive for the test and are reported as a % expression of BCR/ABL over ABL. The sample showing no amplification curve for BCR/ABL target but showing amplification curve for the control are considered as negative for the test and are reported as 0.0% expression. In the absence of amplification curve for the sensitivity of this test is 1 tumor cell in 1,00,000 normal cells.

Recommendations: Quantitative analysis of BCR/ABL transcript level by Real time PCR should ideally be performed before the start of therapy in order to determine the baseline value. The results of this test should always be interpreted in the context of the clinical findings and other haematological & cytogenetic investigations. Samples must be received in the lab within 48 hrs of collection. Specimen processing after 48 hrs of collection may lead to erroneous results due to labile nature of RNA.

Limitations: PCR is a highly sensitive technique; common reasons for paradoxical results are contamination during specimen collection, selection of inappropriate specimens and inherent PCR inhibitors in the specimen.

Note: This test is standardized and validated at SRL Limited

References

1. Br J Hematology (1999); 107:587-99.
2. Haematologica (2000); 85:1248-54.
3. Haematology (2003);279-292

End Of Report

Please visit www.srlworld.com for related Test Information for this accession

Dr. B. R. Das, PhD
Mentor-Molecular Diagnostics

hema CORE™

Your Test Results

Lab No: 10014001
 Patient Name: (N000100001)
 ID No: 10714/10001
 Hospital: (N000100001)
 Address: (N000100001)
 City: (N000100001)
 State: (N000100001)
 Zip: (N000100001)

BCR-ABL1 Quantitative RT-PCR International Units (IU)

BCR-ABL1 (E) (IU)

Received peripheral blood - (E) - collected on 12/03/2010 at 10:00 AM

DOB: 01

Quantitative Real Time PCR

PF35-100x1 52x2 major transcript	Detected
PF35-100x2 minor transcript	Not Detected
PF35-100x3 minor transcript	Not Detected
Observed copies of ABL1	146212
Observed copies of BCR-ABL1	15
BCR-ABL1/ABL1 ratio (%)	0.0003
Conversion Factor for IU	0.58
BCR-ABL1 IU (%)	0.0003

Pattern 100 Historical Results



COMMENTS

1. The hybrid transcript of BCR-ABL1 was quantitated using real-time PCR assay. Signals for BCR-ABL1 were detected in leukocytes of the specimen. Follow-up is recommended, if clinically indicated.



Dr. Shalu Verma Kumar, DDM, Ph.D., Molecular Scientist

[Signature]
Reg. No. 5234

Dr. Raju Katara, Ph.D., Molecular Scientist

[Signature]

CORE DIAGNOSTICS™

Clinical Information

Refrigerated Frozen

Vial ID Barcode



Period: *[Signature]*

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Your Test Results

Case Number: 18014624
Patient Name: Papu Ram Saini (NS0010645)
Age/Sex: 38 Yrs/Male
Patient Location: Jaipur
Hospital Name: Aakriti Labs
Physician Name: Dr. Hemant Malhotra
Date & Time of Accessioning: 03/04/2018 12:00 Hrs
Date & Time of Reporting: 07/04/2018 15:17 Hrs

TEST INFORMATION

Background

1. Chronic myeloid leukaemia (CML), is characterized by the translocation between chromosomes 9 (9q34.1) and 22 (22q11.2). The t(9;22)(q34.1;q11.2) is detected cytogenetically in more than 91-96% of adult CML patients; in 5% of pediatric ALL-B CALLA positive; and 15-30% of adult ALL-B CALLA positive patients. At the molecular level, breaks in the *BCR*, and *ABL1* genes result in the formation of fusion mRNA transcripts.

Assay Description, and Methodology

1. This assay quantifies the Major (p210), Minor (p190), and Micro (p230) transcripts. Its internal reference gene is *ABL1*. It is in accordance with EAC Guidelines, and uses an (EU) CE-IVD approved kit with high sensitivity (>MR4.5). Its IS conversion factor is established in accordance with the guidelines of Europe Against Cancer (EAC), and has been calibrated using WHO International Standards for CML.
2. Total cellular RNA is extracted via silica-membrane-based purification from whole blood or bone marrow collected in EDTA. The assay is an RT-qPCR that uses oligonucleotide hydrolysis principle.
3. Molecular Response (MR) is measured using % *BCR-ABL1* ratio. The formula used is: % *BCR-ABL1* = [No. of copies of *BCR-ABL1* transcripts/No. of copies of control gene transcripts] x 100.
4. For the P210 transcript, this ratio is further normalized to the international scale (IS) and reported as *BCR-ABL1/ABL1* % (IS). The formula used is: % *BCR-ABL1* = [Sum of *BCR-ABL1* copy number/Sum of *ABL1* copy number] * CF x 100. Where * denotes minimum of 10,000 copies.
5. Molecular response is thus expressed and reported as *BCR-ABL1*% on a log scale relative to the standard baseline (100% IS), where 10%, 1%, 0.1%, and 0.0032% correspond to a decrease of 1, 2, 3, and 4.5 logs, respectively, below the standard baseline, i.e. 100% *BCR-ABL1*.
6. The Limit of Detection (LoD) is equal to 2 copies of *BCR-ABL1* transcript. Per Cross et al. (2015), the control gene [*ABL1*] copy numbers for scoring molecular response: MR4.0 = 10,000-31,999 copies of *ABL1*; MR4.5 = 32,000-99,999 copies of *ABL1*; MR5.0 ≥100,000 copies of *ABL1*. This calculation is contingent upon the assay's LOD.
7. Linearity: Major *BCR-ABL1* linearity ranges from 0.0008 to 98 Mbcn NCN, for Minor it ranges from 0.002 to 82 mbcn NCN and Micro *BCR-ABL1* linearity range was 0.005 to 76 jbcn NCN. Here NCN is a normalised copy number obtained from the ratio between *BCR-ABL1* copy number/*ABL1* copy number. Specificity of this assay is 100%.
8. The results should be correlated with clinical data. To monitor response to TKI therapy, NCCN Guidelines Version 2.2017 for CML recommend using this test at diagnosis; Every 3 months after initiating treatment. After *BCR-ABL1* 0.1% - <1% (IS) has been achieved, every 3 months for 2 years and every 3-6 months thereafter; if there is 1-log increase in *BCR-ABL1* transcript levels with MMR, the test should be repeated in 1-3 months.

Disclaimer: The assay is designed to perform the reactions at the specified analytical sensitivity given that the template RNA is not heavily fragmented, and does not contain materials that could inhibit the amplification reaction.



Dr. Shalu Verma Kumar, DVM, Ph.D., Molecular Scientist

Reg. No. 3214

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Dr. Rahul Katara, Ph.D., Molecular Scientist

CÔRE DIAGNOSTICS™

Leukemia CÔRE™ Your Test Results

Case Number: 18014624
Patient Name: Papu Ram Saini (NS0010645)
Age/Sex: 38 Yrs/Male
Patient Location: Jalpur
Hospital Name: Aakriti Labs
Physician Name: Dr. Hemant Malhotra
Date & Time of Accession: 03/04/2018 12:00 Hrs
Date & Time of Reporting: 07/04/2018 15:17 Hrs

REFERENCE

- Druker BJ, et al. IRIS Investigators. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *N Engl J Med.* 2006 Dec 7;355(23):2408-17.
- Branford S et al. Rationale for the recommendations for harmonizing current methodology for detecting BCR-ABL transcripts in patients with chronic myeloid leukaemia. *Leukemia.* 2006 Nov;20(11):1925-30.
- Baccarani M et al., European LeukemiaNet. Evolving concepts in the management of chronic myeloid leukemia: recommendations from an expert panel on behalf of the European LeukemiaNet. *Blood.* 2006 Sep 15;108(6):1809-20.
- Beillard E et al. Evaluation of candidate control genes for diagnosis and residual disease detection in leukemic patients using 'real-time' quantitative reverse-transcriptase polymerase chain reaction (RQ-PCR) - a Europe against cancer program. *Leukemia.* 2003 Dec;17(12):2474-86.
- Cross NC et al. Laboratory recommendations for scoring deep molecular responses following treatment for chronic myeloid leukemia. *Leukemia.* 2015 May;29(5):999-1003.



Dr. Shalu Verma Kumar, DVM, Ph.D., Molecular Scientist

Reg. No. 3214

Dr. Rahul Katara, Ph.D., Molecular Scientist

Page 3 of 4

CÔRE DIAGNOSTICS™

hemaC^{ORE} Your Test Result

Patient ID: _____
 Patient Name: _____
 Age: _____
 Sex: _____
 Hospital Name: _____
 Doctor Name: _____
 Date of Collection: _____
 Date of Report: _____

19060565
 PAPPU SAINI
 40 Yrs/Male
 Jaipur, Rajasthan, India
 Mahatma Gandhi Hospital, Jaipur
 Dr. Hemant Malhotra
 11/06/2019 11:22 Hrs
 13/06/2019 09:20 Hrs

TEST NAME

BCR-ABL1 Quantitative International Scale (IS)

SPECIMEN INFORMATION

Peripheral Blood Collected on 10/06/2019 at 14:30 Hrs

CLINICAL HISTORY

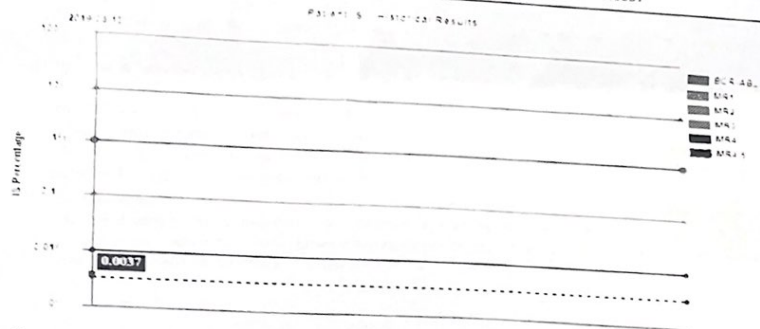
On Glivec 400mg

METHODOLOGY

Real Time Polymerase Chain Reaction (RT-PCR)

TEST RESULT

P210 (b3a2, b2a2) major transcript	Detected
P190 (e1a2) minor transcript	Not detected
P230 (c3a2) micro transcript	Not detected
Observed copies of ABL1	399640
Observed copies of BCR-ABL1	28
BCR-ABL1/ABL1 ratio [%]	0.00690000
Conversion Factor for IS	0.54
BCR-ABL1 IS [%]	0.0037



White blood cell(WBC) count = 5350.00/ μ l; Platelet count = 165000.00/ μ l; Hemoglobin = 12.60 g/dL



Dr. Lata Kini MD Oncopathologist

Lata Kini

Reg No 5641

Dr. Rahul Katara Ph.D. Molecular Scientist

Rahul Katara

Chronic Myeloid Leukemia CORE 40 Hour Test Result

Case ID
Patient Name
Age/Sex
Hospital Location
Hospital Name
Physician Name
Date & Time of Addressing
Date & Time of Reporting

19065665
PAPPU SAINI
40 Yrs/Male
Jaipur, Rajasthan, India
Mahatma Gandhi Hospital, Jaipur
Dr. Hemant Malhotra
11/06/2019 11:22 Hrs
13/06/2019 09:20 Hrs

Disclaimer: The assay is designed to perform the reactions at the specified analytical sensitivity given that the template RNA is not heavily fragmented, and does not contain materials that could inhibit the amplification reaction.

REFERENCES

- Druker BJ, et al. IRIS Investigators. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *N Engl J Med.* 2006 Dec 7;355(23):2408-17.
- Branford S et al. Rationale for the recommendations for harmonizing current methodology for detecting BCR-ABL transcripts in patients with chronic myeloid leukaemia. *Leukemia.* 2006 Nov;20(11):1925-30.
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- Cross NC et al. Laboratory recommendations for scoring deep molecular responses following treatment for chronic myeloid leukemia. *Leukemia.* 2015 May;29(5):999-1003.



Dr. Lata Kini, MD, Oncopathologist

Lata Kini

Reg No. 5641

Page 3 of 4

Dr. Rahul Kataria, Ph.D., Molecular Scientist

Rahul Kataria

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