

Test Description

The MolQ Germline Cancer Predisposition Panel include genes associated with both common and rare hereditary cancers.

Patient Demographic

Name: Ms. Kavita Singh
Sex: Female
Date of Birth/Age: 42 years
Disease: Breast Cancer

Clinician

Clinician Name: Dr Amish Vohra
Medical Facility: Hope Clinic
Pathologist: Not Provided

Specimen

Booking ID: 011908030126
Site: NA
Sample Type: Blood
Date of Collection: 03-08-2019
Date of Booking: 03-08-2019

CLINICAL SYNOPSIS

Kavita Singh, is a case of carcinoma breast. She has been evaluated for pathogenic variations in the genes listed in Appendix 2.

RESULTS

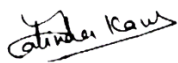
No pathogenic or likely pathogenic variant causative of the reported phenotype are detected

ADDITIONAL FINDINGS: VARIANT(S) OF UNCERTAIN SIGNIFICANCE (VUS) DETECTED

No other variant that warrants to be reported was detected. Variations with high minor allele frequencies which are likely to be benign will be given upon request.

RECOMMENDATIONS

- The sensitivity of NGS based assays to detect large heterozygous deletions/duplications is low and an alternate method is recommended.
- Genetic counselling is advised.



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Head, Molecular Biology & Genomics



Dr. Gulshan Yadav, MD
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APPENDIX 1: TEST METHODOLOGY

Method

Targeted gene sequencing: Selective capture and sequencing of the protein coding regions of the genome/genes is performed. Mutations identified in the exonic regions are generally actionable compared to variations that occur in non-coding regions. Targeted sequencing represents a cost-effective approach to detect variants present in multiple/large genes in an individual.

DNA was used to perform targeted gene capture using a custom capture kit. The libraries were sequenced to mean >80- 100X coverage on Illumina sequencing platform. We follow the GATK best practices framework for identification of variants in the sample using Sentieon (v201808.01)¹. The sequences obtained are aligned to human reference genome (GRCh37/hg19) using Sentieon aligner^{1,2} and analyzed using Sentieon for removing duplicates, recalibration and re-alignment of indels¹. Sentieon haplotypcaller has been used to identify variants which are relevant to the clinical indication. Gene annotation of the variants is performed using VEP program³ against the Ensemble release 91 human gene model⁴. In addition to SNVs and small Indels, copy number variants (CNVs) are detected from targeted sequence data using the ExomeDepth (v1.1.10) method⁵. This algorithm detects rare CNVs based on comparison of the read-depths of the test data with the matched aggregate reference dataset.

Clinically relevant mutations were annotated using published variants in literature and a set of diseases databases - ClinVar, OMIM (updated on 21st November 2018), GWAS, HGMD (v2018.3) and SwissVar⁶⁻⁹. Common variants are filtered based on allele frequency in 1000Genome Phase 3, ExAC (v1.0), gnomAD(v2.1), EVS, dbSNP(v151), 1000 Japanese Genome and reference lab internal Indian population database¹⁰⁻¹⁴. Non-synonymous variants effect is calculated using multiple algorithms such as PolyPhen-2, SIFT, MutationTaster2 and LRT. Only non-synonymous and splice site variants found in the germline cancer predisposition panel genes were used for clinical interpretation. Silent variations that do not result in any change in amino acid in the coding region are not reported.

Total data generated (Gb)	7.79
Total reads aligned (%)	99.99
Reads that passed alignment (%)	93.78
Data Q30 (%)	91.32

^{\$}The classification of the variations is done based on American College of Medical Genetics as described below¹⁵

Variant	A change in a gene. This could be disease causing (pathogenic) or not disease causing (benign).
Pathogenic	A disease causing variation in a gene which can explain the patient's symptoms has been detected. This usually means that a suspected disorder for which testing had been requested has been confirmed.
Likely Pathogenic	A variant which is very likely to contribute to the development of disease however, the scientific evidence is currently insufficient to prove this conclusively. Additional evidence is expected to confirm this assertion of pathogenicity.
Variant of Uncertain Significance	A variant has been detected, but it is difficult to classify it as either pathogenic (disease causing) or benign (non-disease causing) based on current available scientific evidence. Further testing of the patient or family members as recommended by your clinician may be needed. It is probable that their significance can be assessed only with time, subject to availability of scientific evidence.

#The transcript used for clinical reporting generally represents the canonical transcript (according to Ensembl release 87 gene model), which is usually the longest coding transcript with strong/multiple supporting evidence. However, clinically relevant variants annotated in alternate complete coding transcripts could also be reported.

Variants annotated on incomplete and nonsense mediated decay transcripts will not be reported.

#The *in-silico* predictions are based on Variant Effect Predictor, Ensembl release 91 (SIFT version - 5.2.2; PolyPhen - 2.2.2); LRT version - November, 2009 release from dbNSFPv3.1 and Mutation Taster2 based on build NCBI 37 / Ensembl 69.

For any further technical queries please contact contact@molq.in.

DISCLAIMER

- Interpretation of variants in this report is performed to the best knowledge of the laboratory based on the information available at the time of reporting. The classification of variants can change over time and MolQ cannot be held responsible for this. Please feel free to contact MolQ Laboratory (contact@molq.in) in the future to determine if there have been any changes in the classification of any variations. Re-analysis of variants in previously issued reports in light of new evidence

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is not routinely performed, but may be available upon request.

- The sensitivity of this assay to detect large deletions/duplications of more than 10 bp or copy number variations (CNV) is 70-75%. The CNVs detected have to be confirmed by alternate method.
- Due to inherent technology limitations of the assay, not all bases of the exome can be covered by this test. Accordingly, variants in regions of insufficient coverage may not be identified and/or interpreted. Therefore, it is possible that pathogenic variants are present in one or more of the genes analyzed, but have not been detected. The variants not detected by the assay that was performed may impact the phenotype.
- It is also possible that a pathogenic variant is present in a gene that was not selected for analysis and/or interpretation in cases where insufficient phenotypic information is available.
- Genes with pseudogenes, paralog genes and genes with low complexity may have decreased sensitivity and specificity of variant detection and interpretation due to inability of the data and analysis tools to unambiguously determine the origin of the sequence data in such regions.
- The mutations have not been validated/confirmed by Sanger sequencing.
- Incidental or secondary findings (if any) that meet the ACMG guidelines¹⁶ can be given upon request.
- The report shall be generated within turnaround time (TAT), however, such TAT may vary depending upon the complexity of test(s) requested. MolQ Laboratory under no circumstances will be liable for any delay beyond afore mentioned TAT.
- It is hereby clarified that the report(s) generated from the test(s) do not provide any diagnosis or opinion or recommends any cure in any manner. MolQ Laboratory hereby recommends the patient and/or the guardians of the patients, as the case may be, to take assistance of the clinician or a certified physician or doctor, to interpret the report(s) thus generated. MolQ Laboratory hereby disclaims all liability arising in connection with the report(s).
- In a very few cases genetic test may not show the correct results, e.g. because of the quality of the material provided to MolQ Laboratory. In case where any test provided by MolQ Laboratory fails for unforeseeable or unknown reasons that cannot be influenced by MolQ Laboratory in advance, MolQ Laboratory shall not be responsible for the incomplete, potentially misleading or even wrong result of any testing if such could not be recognized by MolQ Laboratory in advance.
- This is a laboratory developed test and the development and the performance characteristics of this test was determined by reference laboratory.

LIMITATIONS

- Genetic testing is an important part of the diagnostic process. However, genetic tests may not always give a definitive answer. In some cases, testing may not identify a genetic variant even though one exists. This may be due to limitations in current medical knowledge or testing technology. Accurate interpretation of test results may require knowing the true biological relationships in a family. Failing to accurately state the biological relationships in {my/my child's} family may result in incorrect interpretation of results, incorrect diagnoses, and/or inconclusive test results.
- Test results are interpreted in the context of clinical findings, family history and other laboratory data. Only variations in genes potentially related to the proband's medical condition are reported. Rare polymorphisms may lead to false negative or positive results. Misinterpretation of results may occur if the information provided is inaccurate or incomplete.
- Specific events like copy number variations, translocations, repeat expansions and chromosomal rearrangements may not be reliably detected with targeted clinical exome sequencing. Variants in untranslated region, promoters and intronic variants are not assessed using this method.
- Genetic testing is highly accurate. Rarely, inaccurate results may occur for various reasons. These reasons include, but are not limited to: mislabelled samples, inaccurate reporting of clinical/medical information, rare technical errors or unusual circumstances such as bone marrow transplantation, blood transfusion; or the presence of change(s) in such a small percentage of cells that may not be detectable by the test (mosaicism).

REFERENCES

1. Freed D. *et al.*, The Sentieon Genomics Tools-A fast and accurate solution to variant calling from next-generation sequence data. *BioRxiv*:115717, 2017.
2. Li H. *et al.*, Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics*, 26(5):589-95, 2010.
3. McLaren W. *et al.*, Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor. *Bioinformatics*, 26(16):2069-70, 2010.
4. Zerbino D. R. *et al.*, Ensembl 2018. *Nucleic Acids Res.*, 46(D1): D754-D761, 2018.
5. Plagnol V. *et al.*, A robust model for read count data in exome sequencing experiments and implications for copy number variant calling. *Bioinformatics*, 28:2747-2754, 2012.

6. Landrum M. J. *et al.*, ClinVar: public archive of interpretations of clinically relevant variants. *Nucleic Acids Res.*, 44(D1):D862-8, 2015.
7. Welter D. *et al.*, The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. *Nucleic Acids Res.*, 42:D1001-1006, 2014.
8. Stenson P. D. *et al.*, The Human Gene Mutation Database: towards a comprehensive repository of inherited mutation data for medical research, genetic diagnosis and next-generation sequencing studies. *Human Genet.*, 136(6): 665-677, 2015.
9. Mottaz A. *et al.*, Easy retrieval of single amino-acid polymorphisms and phenotype information using SwissVar. *Bioinformatics*, 26(6): 851-852, 2010.
10. 1000 Genomes Project Consortium *et al.*, A global reference for human genetic variation. *Nature*, 526(7571): 68-74, 2015.
11. Lek M. *et al.*, Analysis of protein-coding genetic variation in 60,706 humans. *Nature*, 536(7616):285-91, 2016.
12. Exome Variant Server, NHLBI GO Exome Sequencing Project (ESP), Seattle, WA (URL: <http://evs.gs.washington.edu/EVS/>).
13. Nagasaki, M. *et al.*, Rare variant discovery by deep whole-genome sequencing of 1,070 Japanese individuals. *Nat. Commun.*, 6:8018, 2015.
14. Sherry S. T. *et al.*, dbSNP: the NCBI database of genetic variation. *Nucleic Acids Res.*, 29(1):308-11, 2001.
15. Richards S. *et al.*, Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology, *Genet Med.*, 17(5):405-24, 2015.
16. Kalia S.S. *et al.*, Recommendations for reporting of secondary findings in clinical exome and genome sequencing, 2016 update (ACMG SF v2.0): a policy statement of the American College of Medical Genetics and Genomics. *Genet Med.*, 19(2):249-255, 2017.

APPENDIX 2: GENE LIST WITH COVERAGE

Gene	Percentage of coding region covered	Gene	Percentage of coding region covered	Gene	Percentage of coding region covered
<i>AIP</i>	100.00	<i>ALK</i>	100.00	<i>APC</i>	100.00
<i>AR</i>	100.00	<i>ATM</i>	100.00	<i>BAP1</i>	100.00
<i>BARD1</i>	100.00	<i>BLM</i>	100.00	<i>BMPR1A</i>	100.00
<i>BRCA1</i>	100.00	<i>BRCA2</i>	100.00	<i>BRIP1</i>	100.00
<i>BUB1B</i>	100.00	<i>CD82</i>	100.00	<i>CDC73</i>	100.00
<i>CDH1</i>	100.00	<i>CDK4</i>	100.00	<i>CDKN1C</i>	89.35
<i>CDKN2A</i>	100.00	<i>CEBPA</i>	100.00	<i>CEP57</i>	100.00
<i>CHEK2</i>	92.68	<i>CYLD</i>	100.00	<i>DDB2</i>	100.00
<i>DICER1</i>	100.00	<i>DIS3L2</i>	100.00	<i>EGFR</i>	100.00
<i>ELAC2</i>	100.00	<i>ENG</i>	100.00	<i>EPCAM</i>	100.00
<i>ERCC2</i>	100.00	<i>ERCC3</i>	100.00	<i>ERCC4</i>	100.00
<i>ERCC5</i>	100.00	<i>EXT1</i>	100.00	<i>EXT2</i>	100.00
<i>EZH2</i>	100.00	<i>FANCA</i>	100.00	<i>FANCB</i>	100.00
<i>FANCC</i>	100.00	<i>FANCD2</i>	100.00	<i>FANCE</i>	100.00
<i>FANCF</i>	100.00	<i>FANCG</i>	100.00	<i>FANCI</i>	100.00
<i>FANCL</i>	100.00	<i>FANCM</i>	100.00	<i>FH</i>	100.00
<i>FLCN</i>	100.00	<i>GATA2</i>	100.00	<i>GPC3</i>	100.00
<i>HRAS</i>	100.00	<i>KIT</i>	100.00	<i>MAX</i>	100.00
<i>MEN1</i>	100.00	<i>MET</i>	100.00	<i>MLH1</i>	100.00
<i>MLH3</i>	100.00	<i>MRE11A</i>	100.00	<i>MSH2</i>	95.43
<i>MSH3</i>	100.00	<i>MSH6</i>	100.00	<i>MSR1</i>	100.00
<i>MUTYH</i>	100.00	<i>MXI1</i>	100.00	<i>NBN</i>	100.00
<i>NF1</i>	100.00	<i>NF2</i>	100.00	<i>NSD1</i>	99.02
<i>PALB2</i>	100.00	<i>PHOX2B</i>	100.00	<i>PMS1</i>	97.47
<i>PMS2</i>	100.00	<i>PRF1</i>	100.00	<i>PRKAR1A</i>	100.00
<i>PTCH1</i>	100.00	<i>PTEN</i>	100.00	<i>RAD50</i>	100.00
<i>RAD51C</i>	100.00	<i>RAD51D</i>	100.00	<i>RAD54L</i>	100.00
<i>RB1</i>	100.00	<i>RECQL4</i>	100.00	<i>RET</i>	100.00
<i>RHBDF2</i>	100.00	<i>RNASEL</i>	100.00	<i>RUNX1</i>	100.00
<i>SBDS</i>	100.00	<i>SDHAF2</i>	100.00	<i>SDHB</i>	100.00
<i>SDHC</i>	100.00	<i>SDHD</i>	80.17	<i>SLX4</i>	100.00
<i>SMAD4</i>	100.00	<i>SMARCB1</i>	100.00	<i>STK11</i>	100.00
<i>SUFU</i>	100.00	<i>TGFBR2</i>	100.00	<i>TMEM127</i>	100.00
<i>TP53</i>	94.80	<i>TSC1</i>	100.00	<i>TSC2</i>	100.00
<i>VHL</i>	100.00	<i>WRN</i>	100.00	<i>WT1</i>	100.00
<i>XPA</i>	100.00	<i>XPC</i>	100.00		